

Evaluation of droplet dispersion during non-invasive ventilation, oxygen therapy, nebuliser treatment and chest physiotherapy in clinical practice: implications for management of pandemic influenza and other airborne infections

AK Simonds,^{1*} A Hanak,¹ M Chatwin,¹
MJ Morrell,¹ A Hall,² KH Parker,³ JH Siggers³
and RJ Dickinson³

¹Clinical and Academic Unit of Sleep & Breathing, Royal Brompton & Harefield NHS Foundation Trust, London, UK

²Department of Microbiology, Royal Brompton & Harefield NHS Foundation Trust, London, UK

³Department of Bioengineering, Imperial College, London, UK

*Corresponding author

Declared competing interests of authors: AKS has an unrestricted research grant from ResMed Ltd for the investigation of an autotitrating ventilator, which is not related to the present study.



Published October 2010

DOI: 10.3310/hta14460-02

This report should be referenced as follows:

Simonds AK, Hanak A, Chatwin M, Morrell MJ, Hall A, Parker KH, *et al.* Evaluation of droplet dispersion during non-invasive ventilation, oxygen therapy, nebuliser treatment and chest physiotherapy in clinical practice: implications for management of pandemic influenza and other airborne infections. *Health Technol Assess* 2010;**14**(46):131–172.

Health Technology Assessment is indexed and abstracted in *Index Medicus/MEDLINE*, *Excerpta Medica/EMBASE*, *Science Citation Index Expanded (SciSearch®)* and *Current Contents®/Clinical Medicine*.



Abstract

Evaluation of droplet dispersion during non-invasive ventilation, oxygen therapy, nebuliser treatment and chest physiotherapy in clinical practice: implications for management of pandemic influenza and other airborne infections

AK Simonds,^{1*} A Hanak,¹ M Chatwin,¹ MJ Morrell,¹ A Hall,² KH Parker,³ JH Siggers³ and RJ Dickinson³

¹Clinical and Academic Unit of Sleep & Breathing, Royal Brompton & Harefield NHS Foundation Trust, London, UK

²Department of Microbiology, Royal Brompton & Harefield NHS Foundation Trust, London, UK

³Department of Bioengineering, Imperial College, London, UK

*Corresponding author

Background: Influenza viruses are thought to be spread by droplets, but the role of aerosol dissemination is unclear and has not been assessed by previous studies. Oxygen therapy, nebulised medication and ventilatory support are treatments used in clinical practice to treat influenza infection are thought to generate droplets or aerosols.

Objectives: Evaluation of the characteristics of droplet/aerosol dispersion around delivery systems during non-invasive ventilation (NIV), oxygen therapy, nebuliser treatment and chest physiotherapy by measuring droplet size, geographical distribution of droplets, decay in droplets over time after the interventions were discontinued.

Methods: Three groups were studied: (1) normal controls, (2) subjects with coryzal symptoms and (3) adult patients with chronic lung disease who were admitted to hospital with an infective exacerbation. Each group received oxygen therapy, NIV using a vented mask system and a modified circuit with non-vented mask and exhalation filter, and nebulised saline. The patient group had a period of standardised chest physiotherapy treatment. Droplet counts in mean diameter size ranges from 0.3 to > 10 µm were measured with a counter placed adjacent to the face and at a 1-m distance from the subject/patient, at the height of the nose/mouth of an average health-care worker.

Results: NIV using a vented mask produced droplets in the large size range (> 10 µm) in patients

($p=0.042$) and coryzal subjects ($p=0.044$) compared with baseline values, but not in normal controls ($p=0.379$), but this increase in large droplets was not seen using the NIV circuit modification. Chest physiotherapy produced droplets predominantly of > 10 µm ($p=0.003$), which, as with NIV droplet count in the patients, had fallen significantly by 1 m. Oxygen therapy did not increase droplet count in any size range. Nebulised saline delivered droplets in the small- and medium-size aerosol/droplet range, but did not increase large-size droplet count.

Conclusions: NIV and chest physiotherapy are droplet (not aerosol)-generating procedures, producing droplets of > 10 µm in size. Due to their large mass, most fall out on to local surfaces within 1 m. The only device producing an aerosol was the nebuliser and the output profile is consistent with nebuliser characteristics rather than dissemination of large droplets from patients. These findings suggest that health-care workers providing NIV and chest physiotherapy, working within 1 m of an infected patient should have a higher level of respiratory protection, but that infection control measures designed to limit aerosol spread may have less relevance for these procedures. These results may have infection control implications for other airborne infections, such as severe acute respiratory syndrome and tuberculosis, as well as for pandemic influenza infection.



Contents

List of abbreviations	137	NIV using vented mask	147
Executive summary	139	Modified NIV	147
1 Introduction	141	Oxygen therapy	148
2 Methods	143	Nebuliser therapy	148
Trial design	143	4 Discussion	155
Droplet visualisation	143	Limitations	155
Equipment	144	5 Conclusions	157
Interventions	144	Acknowledgements	159
Study area/baseline readings	145	References	161
Analysis	145	Appendix I Protocol	163
3 Results	147		
Physiotherapy	147		



List of abbreviations

AGP	aerosol-generating procedure	O ₂	oxygen therapy
CI	confidence interval	PaCO ₂	partial pressure of arterial carbon dioxide
COPD	chronic obstructive pulmonary disease	PaO ₂	partial pressure of arterial oxygen
CPAP	continuous positive airway pressure	PPE	protective personal equipment
DH	Department of Health	RR	relative risk
EPAP	expiratory positive airway pressure	SaO ₂	arterial oxygen saturation
IPAP	inspiratory positive airway pressure	SARS	severe acute respiratory syndrome
mod NIV	modified NIV circuit with exhalation filter	SD	standard deviation
NIV	non-invasive ventilation	TcCO ₂	transcutaneous partial pressure of carbon dioxide
		WHO	World Health Organization

All abbreviations that have been used in this report are listed here unless the abbreviation is well known (e.g. NHS), or it has been used only once, or it is a non-standard abbreviation used only in figures/tables/appendices, in which case the abbreviation is defined in the figure legend or in the notes at the end of the table.



Executive summary

Background

Influenza viruses are thought to be spread by droplets, but the role of aerosol dissemination (defined as droplet size range $< 5 \mu\text{m}$) is unclear. A subgroup of patients, often with underlying chronic disorders or risk factors, such as pregnancy or immunosuppression, can develop pneumonia/respiratory insufficiency with H1N1 swine flu or other influenzal infection requiring treatment by oxygen therapy (O₂), nebulised medication and ventilatory support. These therapies are thought to generate droplets or aerosols, and in the severe acute respiratory syndrome (SARS) outbreak were associated with an increased incidence of SARS in health-care workers and higher risk of superspreading events in hospital wards. Non-invasive ventilation (NIV) is unlikely to be effective in rapidly progressive acute lung injury, but may have a role in chronic patients in whom influenza has caused an infective exacerbation, and its use may reduce pressure on intensive care beds. Previous studies have not assessed droplet or aerosol generation during respiratory support interventions in clinical practice.

Objectives

We evaluated the characteristics of droplet/aerosol dispersion around delivery systems during NIV, O₂, nebuliser treatment and chest physiotherapy by measuring droplet size, geographical distribution of droplets, decay in droplets over time after the interventions were discontinued, and the impact of modification of the NIV circuit in clinical practice.

Methods

Three groups were studied: (1) normal control subjects, (2) subjects with coryzal symptoms and (3) adult patients with chronic lung disease who were admitted to hospital with an infective exacerbation.

Each group received O₂, NIV using a vented mask system and a modified circuit with non-vented mask and exhalation filter, and nebulised saline. The patient group had a period of standardised chest physiotherapy treatment. Droplet counts in

mean diameter size ranges from 0.3 to $> 10 \mu\text{m}$ were measured with a counter placed adjacent to the face (D1) and at 1-m distance (D2) from subject/patient at the height of the nose/mouth of an average health-care worker.

Results

Non-invasive ventilation using a vented mask produced droplets in the large size range ($> 10 \mu\text{m}$) in patients ($p = 0.042$) and coryzal subjects ($p = 0.044$) compared with baseline values, but not in normal controls ($p = 0.379$). This increase in large droplets was not seen using the NIV circuit modification. Chest physiotherapy produced droplets predominantly of $> 10 \mu\text{m}$ ($p = 0.003$), which, as with NIV droplet count in the patients, had fallen significantly by 1 m. O₂ did not increase droplet count in any size range. Nebulised saline delivered droplets in the small- and medium-size aerosol/droplet range in keeping with the specified performance characteristics of the device but did not increase large-size droplet count. Preliminary analysis suggests that droplet counts fall to within a baseline range within 20–40 minutes of discontinuing the NIV and chest physiotherapy.

Conclusions

Non-invasive ventilation and chest physiotherapy are droplet (not aerosol)-generating procedures, producing droplets of $> 10 \mu\text{m}$ in size. Due to their large mass, most fall out on to local surfaces within 1 m. The only device producing an aerosol was the nebuliser and the output profile is consistent with nebuliser characteristics rather than dissemination of large droplets from patients. These findings suggest that health-care workers providing NIV and chest physiotherapy working within 1 m of an infected patient should have a higher level of respiratory protection, but that infection control measures designed to limit aerosol spread, for example negative-pressure rooms, may have less relevance. The results may have infection control implications for other airborne infections, such as SARS and tuberculosis, as well as for pandemic influenza infection.

Chapter I

Introduction

Respiratory viral infections, such as influenza, are spread by droplets, an aerosol of infected material or by direct or indirect contact with contaminated surfaces. The mode of transmission and the factors influencing this are important, as they have key implications for infection control in patients and staff, and therefore pandemic planning. Droplets in the respirable range (around 5 µm) may play a significant part in transmission,¹ but the role of aerosols has been questioned² and there are few studies quantifying viral load in droplets or aerosols. An observational study³ of influenza A and influenza B in exhaled breath showed viral RNA in one-third of infected patients, and 99% of particles had a diameter of < 5 µm when sampled during tidal breathing.

While some individuals recover from seasonal or H1N1 influenza, having experienced minimal symptoms, a subgroup of high-risk patients may develop complications, including respiratory failure,^{4,5} and, in new more pathogenic strains, such as H5N1, respiratory insufficiency may occur in more than 50% of those affected. These patients are managed with antiviral therapy and antibiotics for secondary bacterial pneumonia, but the mainstay of management is supportive respiratory care, which includes high-flow oxygen therapy (O₂) for hypoxaemic patients, and ventilatory support for those with ventilatory insufficiency.^{6,7} Adjunctive therapy can include nebulised bronchodilator for patients with underlying asthma or chronic obstructive pulmonary disease (COPD), and physiotherapy is used to facilitate secretion clearance for those in whom influenza has precipitated an infective exacerbation of chronic lung disease, such as COPD, bronchiectasis or cystic fibrosis.

Coughing and sneezing patients can shed relatively large particles (> 10 µm) that travel short distances and may contaminate the bedside environment. Smaller droplets or aerosols will remain airborne for longer periods and disseminate over greater distances.¹ The definition of an aerosol varies but most authorities characterise this as consisting of droplets of < 5 µm. Some medical procedures have been termed ‘aerosol generating’, as the common feature is that they are associated with high or

augmented inspiratory and expiratory tidal flows, which may increase viral dissemination but this classification is based on assumptions rather than systematic evidence. The list of aerosol-generating procedures (AGPs) differs a little from country to country but in Department of Health (DH) guidance^{7,8} these include bronchoscopy, intubation of the airway and invasive ventilation manoeuvres, such as open suctioning, cardiopulmonary resuscitation, non-invasive ventilation (NIV) and continuous positive airway pressure (CPAP) therapy, high-frequency oscillation ventilation, and induction of sputum. Certain other procedures, such as delivery of nebulised medication therapy and high-flow O₂ are considered to be possible aerosol generators, but a lesser infective risk.⁸ There is an association between some of these AGPs and an increased incidence of severe acute respiratory syndrome (SARS) in health-care workers^{9–11} and the risk of superspreading events on wards.¹² This has implications for the safe care of patients and risk management for nurses, doctors, physiotherapists and other health-care workers, and has provoked an ethical debate on the duty of care of health-care staff in pandemics.^{13,14}

Much of the evidence for the link between AGPs and increased transmission of respiratory viral infection was generated during the SARS epidemic. In Toronto and Singapore, health-care workers constituted approximately 20% of critically ill cases. Infection rates were higher in doctors and nurses carrying out endotracheal intubation [relative risk (RR) 13.29, 95% confidence interval (CI) 2.99 to 59.04, $p = 0.03$], while nurses caring for SARS patients receiving NIV may have been at increased risk (RR 2.23), but this finding did not reach significance (95% CI 0.25 to 21.76, $p = 0.5$).⁹ In a case-control study of dissemination of SARS from an index case to other patients on the same ward, Yu *et al.*¹² showed an increased risk associated with the index patient requiring O₂ or bilevel NIV. Case reports^{15,16} have also linked transmission of infection to nebuliser use in the index patient. However, patient variables are also likely to be important, as sicker patients who may have a higher viral load are more likely to require O₂ and ventilatory support, and those with underlying asthma who require nebuliser therapy

may cough more due to airway hyper-reactivity. For these reasons specific infection control precautions have been introduced for unavoidable AGPs and these include use of high-efficiency FFP3 (or N95) masks, eye protection, gowns, aprons and gloves.⁸ Guidelines also suggest that AGPs should only be used if necessary, and controversy has arisen over the role of NIV.^{17,18} Its use is recommended with appropriate precautions in some national guidelines,⁷ but not in other guidelines, and NIV use is cautioned against by some authorities.¹⁹⁻²¹

Non-invasive ventilation (NIV) and CPAP are unlikely to have a role in acute lung injury caused by influenza or in secondary bacterial pneumonia, or in patients with multisystem failure.¹⁷ However, NIV was used successfully in some SARS cases,^{22,23} and as indicated in DH guidelines,²⁴ there is potential for NIV to reduce the need for intubation in influenza pneumonia in those with chronic respiratory disease,²⁵ to facilitate extubation, and to widen the provision of ventilatory support outside the intensive care unit. It may also be used as a ceiling of ventilatory care in patients with COPD, congestive cardiac failure and other serious comorbidities, and to palliate symptoms in those with end-stage disease in whom ICU admission is not indicated. These indications should be set against the risks of droplet dissemination during the delivery of NIV – yet at present those risks have not been quantified.

It is also important to note that there are problems in interpreting the evidence of transmission of infection during SARS. This is because transmissibility could have been increased by an inadequate use of protective personal equipment (PPE) in early cases;^{11,26} NIV equipment has evolved since 2003–4, and there have been subsequent experimental studies that have investigated air flows around oxygen masks and during NIV.²⁷⁻³⁰ These studies used human simulator models or normal subjects mimicking respiratory distress. Hui *et al.*^{28,31} have carried out a series of experimental studies analysing particle spread from NIV and oxygen masks,³² using smoke particles as a proxy of droplets in expired air. However, human simulators may not closely reflect the behaviour of sick patients, and smoke particles are considerably smaller (< 1 µm) than droplets generated by coughing and sneezing (range

5 to > 10 µm). Therefore, the behaviour of smoke particles may not accurately represent droplet dispersion. Other workers have used a Schlieren optical visualisation technique³³ to demonstrate exhaled air flows in normal subjects when coughing with and without masks. These provide useful information on expiratory flow profiles but none of the investigations has been carried out using the range of common clinical interventions defined as AGPs, analysed droplet size or studied patients with respiratory infections.

This background therefore provided the rationale of this study, the aim of which was to investigate droplet dispersion during O₂, NIV and nebuliser treatment in patients with coryzal symptoms, patients with an infective exacerbation of chronic lung disease and a control group of normal subjects, to inform safe use. We reasoned that patients with a chronic exacerbation of lung disease or a coryzal infection would generate droplets regardless of the aetiology of the infection, therefore we did not specify that the infection had to be due to H1N1 or any other subtype of influenza A or influenza B. We sought to:

1. determine droplet size and concentration
2. determine geographical distribution of droplets
3. compare and contrast droplets generated during different interventions
4. examine whether modifications of treatment delivery affect droplet dissemination
5. estimate droplet decay after the intervention had ceased.

Although not classified as an AGP, we added an analysis of droplet counts and dispersion during a standardised session of chest physiotherapy in the chronic respiratory patients. This was because there was a high level of concern by physiotherapists that droplet dissemination would be considerable, thus putting these health-care workers at risk. In addition, in 40 patients admitted to the respiratory wards at the Royal Brompton Hospital NHS Foundation Trust with suspected swine flu in the first and second wave of H1N1 in 2009, all had underlying respiratory disease (predominantly cystic fibrosis and asthma) or neuromuscular disease, and required chest physiotherapy as part of their clinical management.

Chapter 2

Methods

Trial design

This was an observational trial carried out in a standard single-bedside room on a respiratory ward at the Royal Brompton & Harefield NHS Foundation Trust. The study was approved by Brompton, Harefield and NHLI Research Ethics Committee (ref. no. 09/H0708/58).

Normal subjects

Normal subjects were recruited from a departmental database of normal people aged 18 years and above. Individuals with a current illness or underlying condition were excluded.

Coryzal patients

To fulfil entry criteria these patients were individuals, aged 18 years and above, who were previously well with no underlying health condition but, within the previous 24–48 hours, had developed a pyrexia or history of pyrexia and any two of the following flu-like symptoms: sore throat, muscle aches and pains, cough and/or headache.

Patients

We recruited patients with an acute infective exacerbation of chronic respiratory disease requiring admission to a respiratory ward. Inclusion criteria: aged 18 years and above, clinically confirmed infective exacerbation and with an underlying diagnosis of asthma, cystic fibrosis, COPD, bronchiectasis or chest wall disease for which O₂ and/or NIV was clinically indicated. Exclusion criteria: haemodynamic instability, partial pressure of arterial oxygen (PaO₂) < 7.4 kPa, partial pressure of arterial carbon dioxide (PaCO₂) > 7.5 kPa, pH < 7.34 on oxygen/NIV therapy, cognitive inability such that patient was unable to understand information sheet or that the patient was unable to breathe spontaneously for more than 4 hours.

Droplet visualisation

Droplets were detected using an optical particle sizer (Aerotrak 8220, TSI Instruments Ltd, High Wycombe, UK), which counts particles in the range 0.3 to > 10 µm within ranges of 0.3–0.5, 0.5–1.0, 1.0–3.0, 3.0–5.0, 5.0–10.0 and > 10.0 µm, with a counting efficiency of 50% ± 10% at 0.3 µm and 100% ± 10% at 0.45 µm and greater. Particles or droplets are measured in size and concentration per cubic metre by detecting the light scattered from individual droplets as they are drawn through a focused laser beam. The intensity of scattered light is a composite function of the diameter, shape and refractive index of the droplet, as well as the light wavelength and the geometry of the optical detector. A photodetector within the Aerotrak measures the amount of light each droplet scatters and records a count for each size range, for example 0.3–0.5 µm, 5–10 µm, etc. The two Aerotrak devices were calibrated before and after the series of normal subject, coryzal and patient study runs, using polystyrene latex spheres made to particle standards in each size band from 0.3–10 µm. The count efficiency of the device for droplets of 0.45 µm and larger was 100% ± 10%, and at 0.3 µm it was 50% ± 10%. The baseline zero count assurance test using a HEPA filter was passed at a count of < 1 particle per 5 minutes at a 95% confidence level in accordance with ISO (International Organisation for Standardisation) 21501–4. Sampling flow rates of both Aerotrak devices were within 5% of tolerance when calibrated before and after the study runs.

Droplet sampling was carried out over 30 seconds, at 5-minute intervals during baseline periods, and treatment interventions with an Aerotrak detector placed at two sampling points: D1, adjacent (within 20 cm) to patient/subject mouth or treatment mask/interface, and D2, 1.0 m from subject/patient at 45 degrees in the lateral plane. The position D2 was chosen to represent a typical location of a health-care worker providing assistance to the patient. Each Aerotrak counter was zeroed using a HEPA filter before each study run. To maintain

accuracy and reproducibility of measurements, the Aerotrak at D1 was placed in a fixed position on a bed table and the Aerotrak at 1.0 m was mounted on a tripod, adjusted to a height of approximately 1.52 m (5 ft) from the floor, which is equivalent to the height of the nose/mouth of an average-sized health-care worker.

Equipment

Non-invasive ventilation (NIV) was provided using a VPAP ST III (ResMed UK Ltd, Abingdon, UK) bilevel positive pressure ventilator set in spontaneous timed mode. NIV was delivered (1) using a vented full-face mask that was sized to subject (ResMed vented hospital-use face mask) or (2) using a modified circuit. The modified circuit consisted of non-vented full-face mask (ResMed non-vented hospital face mask) and a viral/bacterial filter (Intersurgical filter 1944) placed between the mask and an expiratory leak so that exhalate was filtered [modified NIV (mod NIV)]. The ventilator was started once the mask was secured on the face. In the normal subjects and the coryzal group the ventilator settings were: inspiratory positive airway pressure (IPAP) 20 cmH₂O, expiratory positive airway pressure (EPAP) 5 cmH₂O, with back-up rate of 15 per minute. In patients the IPAP, EPAP and back-up rate were set at clinically required levels, with oxygen entrained into the NIV circuit as clinically indicated to maintain arterial oxygen saturation of > 90%. We used a standard

jet nebuliser with compressor (Actineb, Clement Clark International Ltd, Harlow, UK), which was designed to generate a droplet profile of mass mean diameter 3.3 µm, with 72% droplets < 5 µm, at average flow rate of 7 l/minute. In each nebuliser intervention this delivered 4 ml of normal saline to the normal subjects, coryzal subjects and the patients.

Interventions

Normal controls and coryzal patients

On arrival in the side room, subjects were seated in a semirecumbent position on the bed. The Aerotrak counters were aligned to the subject as described, and baseline readings of droplet counts at the two sampling positions D1 and D2 were obtained over 40 minutes, sampling at 5-minute intervals. Subjects were then asked to do a series of spontaneous coughs both without and with a surgical mask. They then received O₂ via a 60% Ventimask for 20 minutes, then NIV delivered through the non-vented hospital full-face mask (ResMed) using the modified filtered circuit for 20 minutes, then NIV via standard circuit with a vented mask for 20 minutes, and, finally, 4 ml of nebulised normal saline via the mouthpiece. Between interventions there were periods of 40 minutes to allow background droplet counts to fall to baseline levels (*Figure 1*).

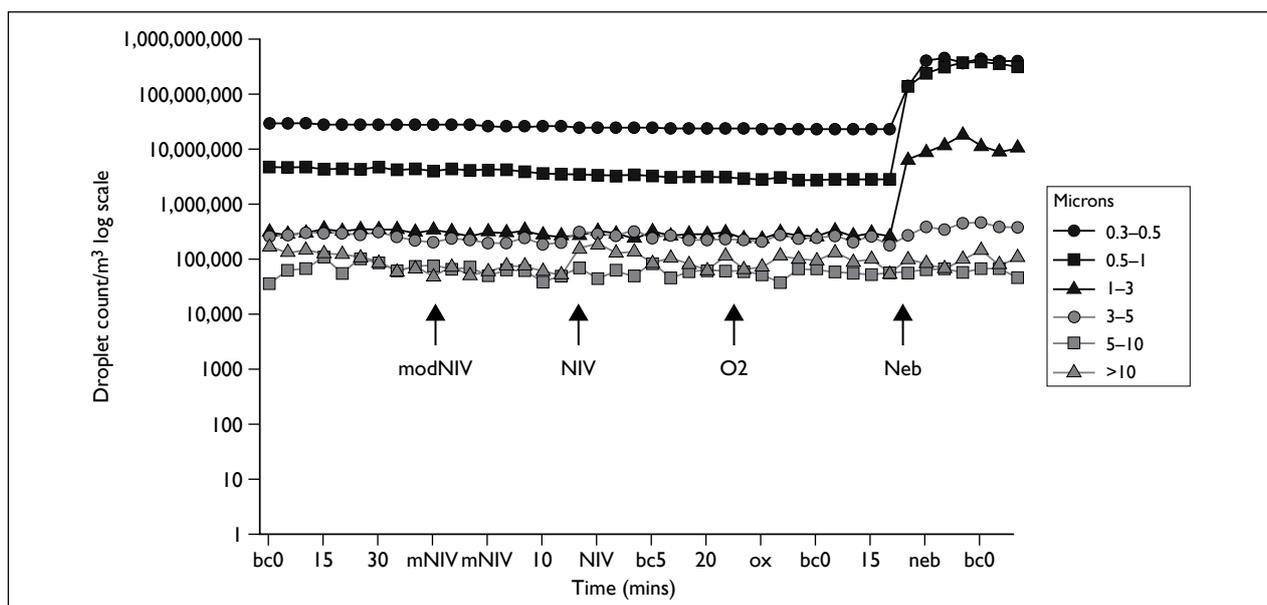


FIGURE 1 Example of coryzal subject experimental run. bc, baseline count; mNIV, modified NIV; neb, nebuliser; ox, oxygen therapy.

Patients

Measurements were carried out as above but with the following differences. All patients had arterial oxygen saturation (SaO_2), heart rate and transcutaneous PCO_2 monitored throughout the experimental interventions. After baseline measurements patients received 24% oxygen via Venturi mask for 20 minutes. Those who were using NIV received 20 minutes of ventilatory support at their current clinically indicated IPAP and EPAP settings, with oxygen entrained to maintain saturation to $> 90\%$, first with the modified circuit and non-vented mask (2) and secondly with the vented face mask (1). The patients also underwent a standardised session of chest physiotherapy over 10 minutes. This consisted of cycles of deep breathing with percussion or shaking to loosen any secretions, followed by an assisted cough initiated manually, augmented by the physiotherapist performing inward and upwards pressure on the lower thorax to aid expectoration, after which the patient rested and cycles were repeated for 10 minutes. Throughout the study only two physiotherapists performed the physiotherapy in order to standardise the techniques as far as possible. Intervals of 40 minutes between interventions were added as in normal subjects and coryzal patients to re-establish baseline droplet levels (Figure 2).

Study area/baseline readings

A standard ward side room, of width and length 3.37×3.37 m and height 2.84 m, was used for all experiments. There was no external window or external ventilation system. Disturbances in the room were minimised by keeping the door shut throughout and allowing one investigator to be present. The investigator wore a surgical mask throughout, and provided the physiotherapy. The experiments were usually undertaken in two runs, in the morning and afternoon of the same day, each lasting approximately 2.5 hours. This length of time was needed in order to allow 20–40 minutes between consecutive interventions so that baseline droplet counts could be restored. Patients and subjects rested in a position of comfort in the bed throughout the interventions, and the position was not changed between the interventions in order to maintain D1 and D2 distances.

Analysis

There are few previous data on droplets generated by respiratory interventions on which to base the sample size. We reasoned that if infection is predominantly transmitted by coughing and sneezing then an increase in droplet count caused

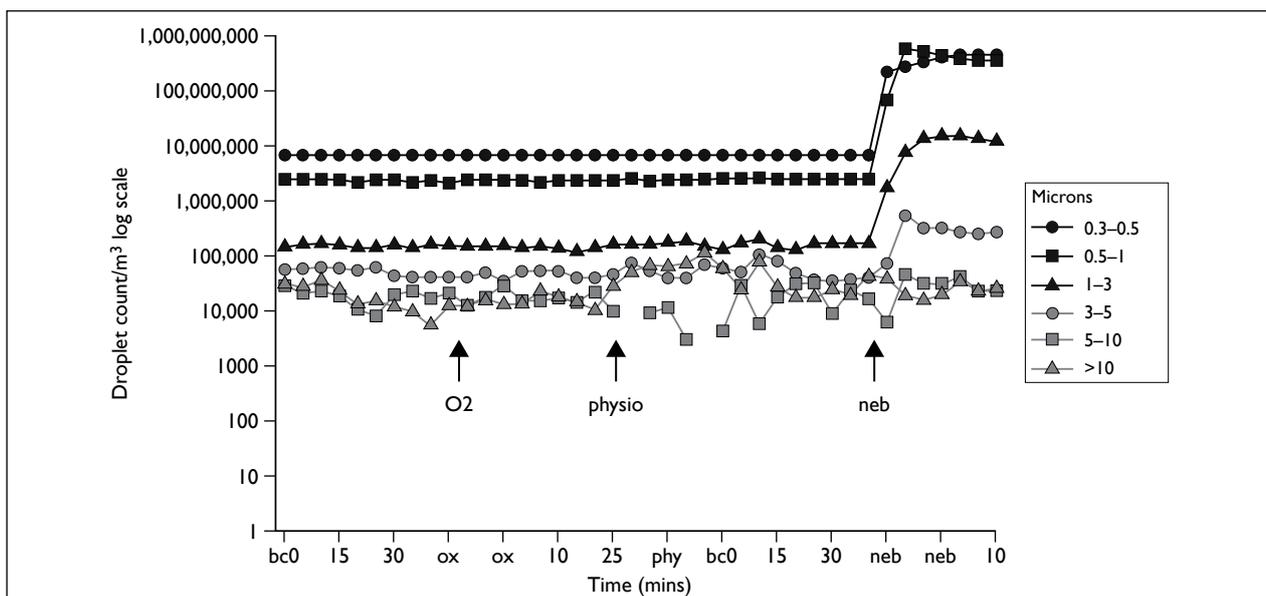


FIGURE 2 Example of patient experimental run. bc, baseline count; neb, nebuliser; ox, oxygen therapy.

by interventions, equivalent to the increase seen from tidal breathing to spontaneous cough, would be clinically meaningful. Pilot studies suggested that a doubling in droplet size would occur in the ranges of 5–10 μm and > 10 μm , with possibly a greater increase in the aerosol range. A doubling in count of 5–10 μm or > 10 μm from a mean of 900 in 5–10 or > 10 μm range to 1800 [standard deviation (SD) 100], with a false-positive rate of 0.05 and 80% power, suggested that very small groups would be needed. We increased the patient group size to account for the possibility that counts and variability might be higher in patients and therefore aimed to study a minimum of 10 normal subjects, 10 with coryzal symptoms and 20 patients.

For each of the five interventions, droplet sampling was carried out on four occasions at 5-minute intervals before treatment, and on four occasions during the intervention, and these values were averaged to give N_{pre} and N_{post} . As we were interested in the relative change due to the intervention rather than absolute values in each subject, this difference was normalised by the average of the four control samples taken before the intervention to give the normalised difference Δ or D $(N_{\text{post}} - N_{\text{pre}}) / N_{\text{pre}}$ was calculated. The significance of this normalised difference was calculated using the two-sided Student's *t*-test.

Chapter 3

Results

In total, 44 subjects and patients were studied: 12 normal controls, 11 with coryzal symptoms and 21 patients. Subject and patient characteristics are given in *Tables 1–3*. The patients had a range of chronic lung conditions and all had been admitted because of an acute infective exacerbation. None of the patients or coryzal individuals had an H1N1 infection. All normal subjects and 10 of the coryzal subjects completed the 60% O₂, NIV, mod NIV and nebuliser therapy. One coryzal patient completed all interventions except NIV modes, as these provoked claustrophobia. All patients received physiotherapy, but normal subjects or coryzal patients did not; all patients received 24% O₂ via Ventimask and nebuliser therapy. Eight patients received NIV and mod NIV, as this was indicated to manage hypercapnic respiratory failure. A total of 19 coryzal subjects and patients therefore underwent the NIV and mod NIV interventions.

Physiotherapy

In the patients there was an increase in > 10- μ m droplets at D1, but this has fallen at 1 m (D2) ($p < 0.003$) (*Figure 3*). There was no increase in the other droplet ranges.

NIV using vented mask

The mean difference increased in the coryzal and patient group in the > 10- μ m range at D1, but not in the normal controls, and this count was elevated at D2 in 3–5 μ m, 5–10 μ m and > 10- μ m ranges at D2 in the coryzal subjects.

Modified NIV

Using the circuit modification the mean difference was not significantly different from baseline values

TABLE 1 Normal subjects: age and trial interventions

Normal subject no.	Age (years)	Trial interventions
1	38	O ₂ , NIV, mod NIV, Neb
2	35	O ₂ , NIV, mod NIV, Neb
3	52	O ₂ , NIV, mod NIV, Neb
4	24	O ₂ , NIV, mod NIV, Neb
5	32	O ₂ , NIV, mod NIV, Neb
6	52	O ₂ , NIV, mod NIV, Neb
7	32	O ₂ , NIV, mod NIV, Neb
8	28	O ₂ , NIV, mod NIV, Neb
9	25	O ₂ , NIV, mod NIV, Neb
10	24	O ₂ , NIV, mod NIV, Neb
11	28	O ₂ , NIV, mod NIV, Neb
12	34	O ₂ , NIV, mod NIV, Neb
Mean (SD)	33.7 (9.6)	

mod NIV, modified non-invasive ventilation circuit; Neb, nebulised saline; NIV, non-invasive ventilation; O₂, oxygen therapy 60%; SD, standard deviation.

TABLE 2 Coryzal group: age and interventions

Coryzal patient no.	Age (years)	Trial interventions
1	30	O ₂ , Neb ^a
2	24	O ₂ , NIV, mod NIV, Neb
3	32	O ₂ , NIV, mod NIV, Neb
4	45	O ₂ , NIV, mod NIV, Neb
5	28	O ₂ , NIV, mod NIV, Neb
6	37	O ₂ , NIV, mod NIV, Neb
7	25	O ₂ , NIV, mod NIV, Neb
8	38	O ₂ , NIV, mod NIV, Neb
9	24	O ₂ , NIV, mod NIV, Neb
10	28	O ₂ , NIV, mod NIV, Neb
11	30	O ₂ , NIV, mod NIV, Neb
Mean (SD)	31 (6.6)	

mod NIV, modified NIV circuit; Neb, nebulised saline; NIV, non-invasive ventilation; O₂, oxygen therapy 60%; SD, standard deviation.
 a Coryzal patient no. 1 did not complete NIV and mod NIV interventions due to claustrophobia.

TABLE 3 Patient age, diagnosis and interventions

Patient	Age (years)	Diagnosis	Indication for admission	Study interventions
1	74	Bronchiectasis	Infective exacerbation	O2, NIV, mod NIV, Neb, Physio
2	55	COPD	Infective exacerbation	O2, NIV, mod NIV, Neb, Physio
3	37	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
4	34	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
5	18	Bronchiectasis	Infective exacerbation	O2, Physio, Neb
6	27	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
7	29	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
8	58	Bronchiectasis	Infective exacerbation	O2, Physio, Neb
9	62	Bronchiectasis	Infective exacerbation	O2, Physio, Neb
10	20	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
11	25	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
12	64	Obesity hypoventilation syndrome	Chest infection	O2, NIV, mod NIV, Neb, Physio
13	48	Bronchopulmonary aspergillosis	Infective exacerbation	O2, NIV, mod NIV, Neb, Physio
14	39	Scoliosis	Chest infection	O2, NIV, mod NIV, Neb, Physio
15	80	COPD	Infective exacerbation	O2, NIV, mod NIV, Neb, Physio
16	59	Asthma	Infective exacerbation	O2, NIV, mod NIV, Neb, Physio
17	58	Bronchiectasis	Infective exacerbation	O2, NIV, mod NIV, Neb, Physio
18	24	Cystic fibrosis	Infective exacerbation	O2, NIV, mod NIV, Neb, Physio
19	44	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
20	27	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
21	18	Cystic fibrosis	Infective exacerbation	O2, Physio, Neb
Mean (SD)	42.8 (19.1)			

COPD, chronic obstructive pulmonary disease; mod NIV, modified NIV circuit; Neb, nebulised saline; NIV, non-invasive ventilation; O2, oxygen therapy 24%; Physio, chest physiotherapy; SD, standard deviation.

on NIV in any group at D1 or D2, indicating that droplet count was significantly reduced compared with standard NIV with vented mask (*Figure 4*).

Oxygen therapy

In normal controls, the coryzal group and in patients no significant increase in droplets in aerosol or large droplet range was seen either at D1 or D2 (*Figure 5*).

Nebuliser therapy

In all groups there was a significant increase across all droplet size ranges on therapy at D1 and D2 in normal subjects. In coryzal subjects and patients there were increases in the 0.3–0.5, 0.5–1, 1–3 and 3.5-µm aerosol ranges both at D1 and D2, but no significant mean difference in the larger droplet ranges of 5–10 and > 10 µm (*Figure 6*).

Mean differences and *p*-values for all interventions in each group are given in *Table 4*.

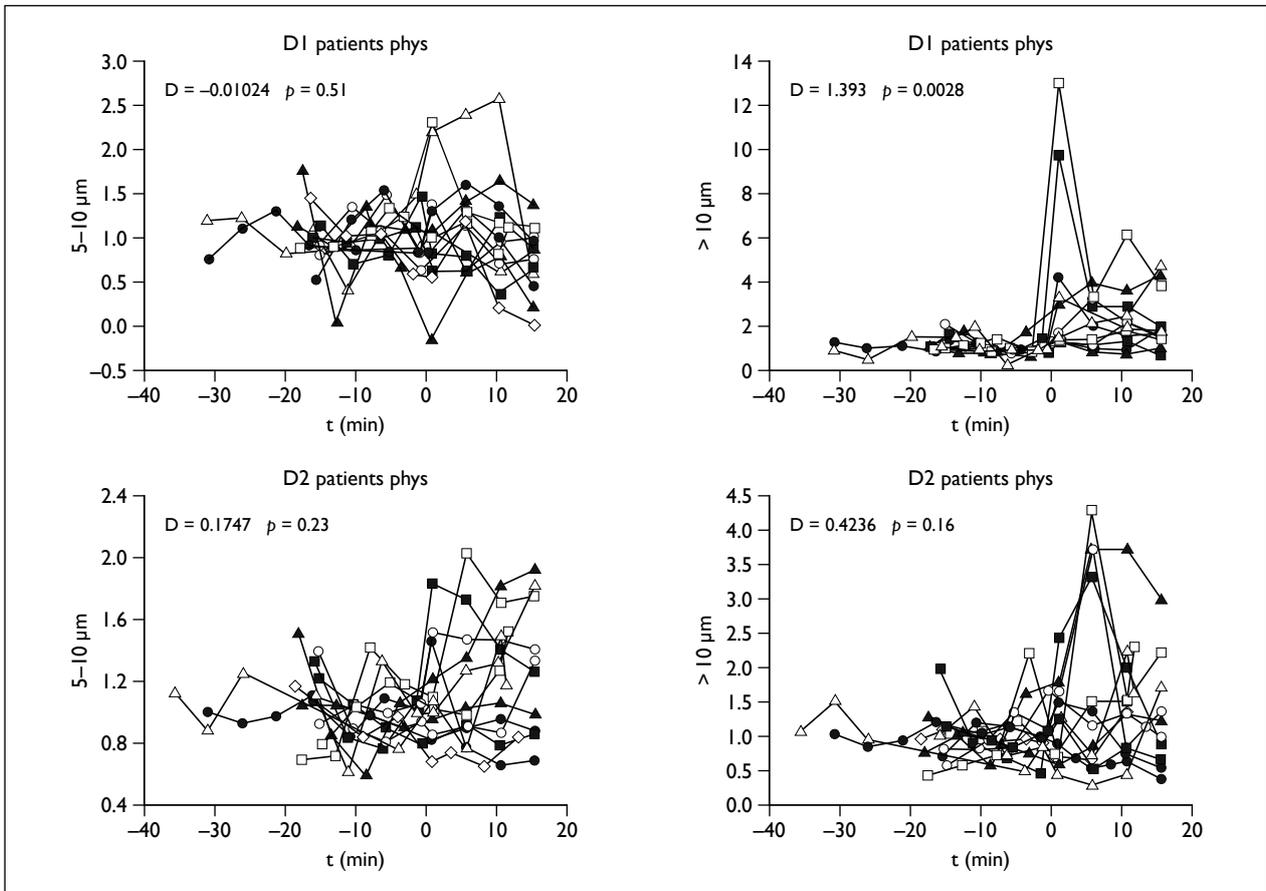


FIGURE 3 Physiotherapy results: droplet size > 10 µm at D1 and D2 in patients.

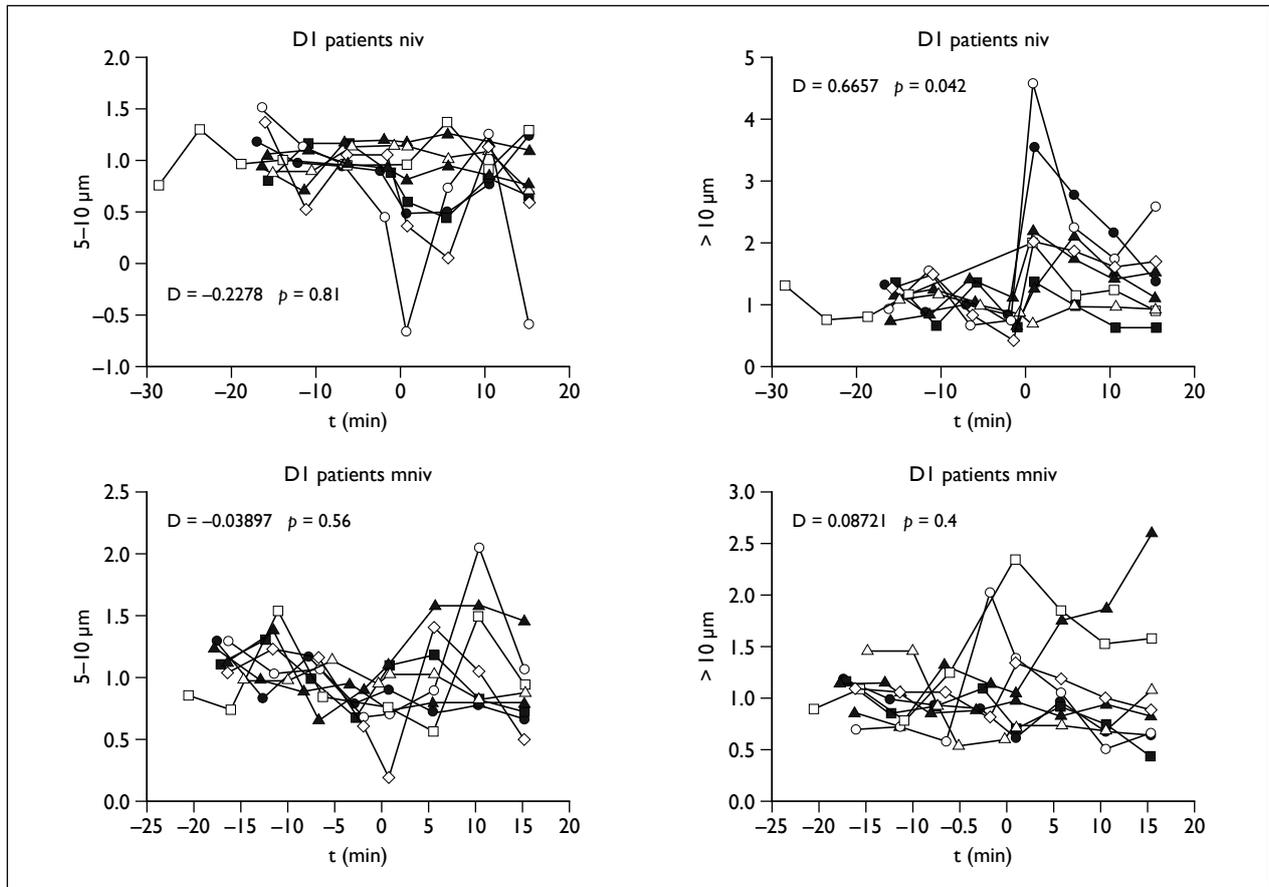


FIGURE 4 Non-invasive ventilation circuit (NIV) (top row) and modified NIV results (bottom row) in patients at DI in ranges 5–10 μm and > 10 μm .

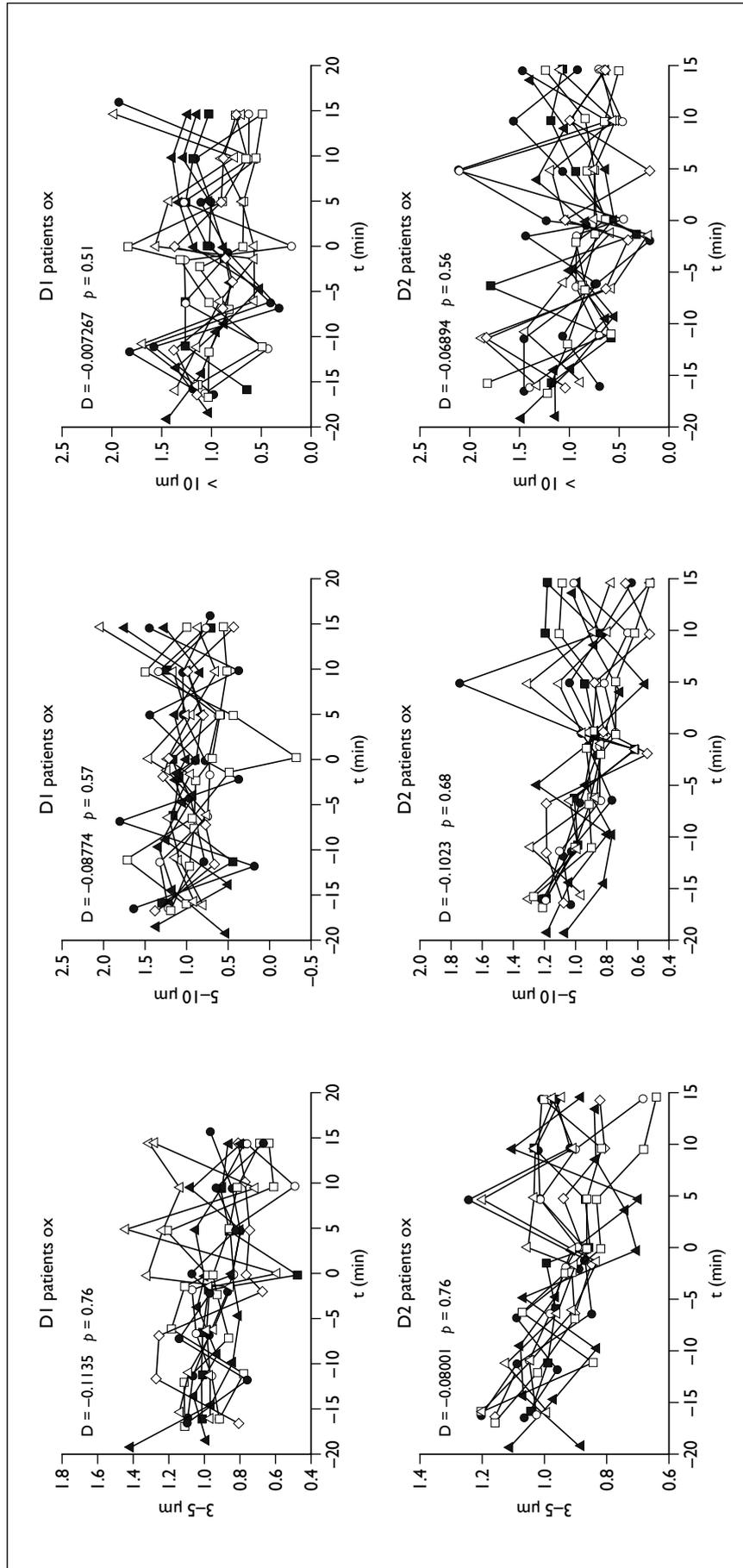


FIGURE 5 Oxygen results in patients at D1 and D2 in droplet ranges 3–5, 5–10 and > 10 μm.

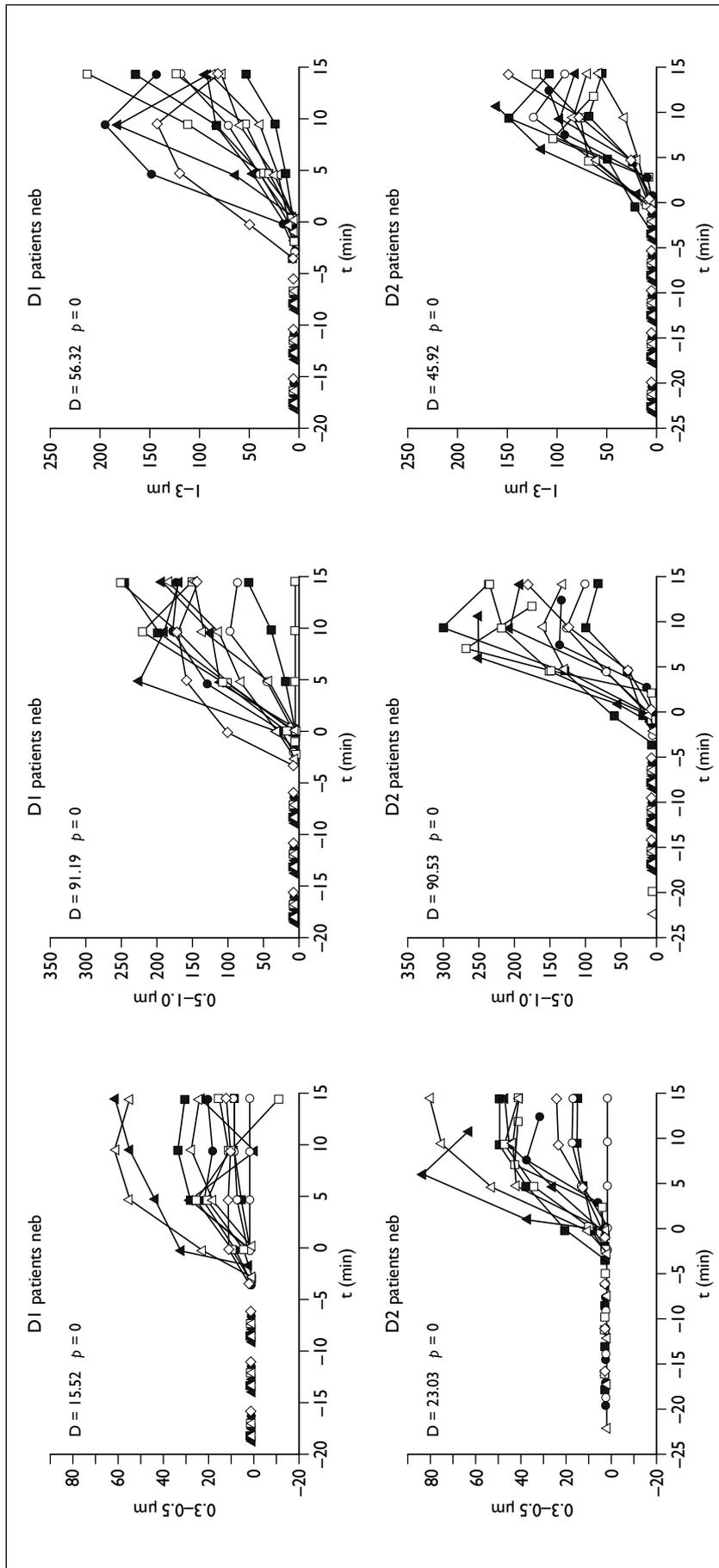


FIGURE 6 Nebuliser results in patients at D1 and D2 in droplet ranges 0.3–0.5 to 1–3 μm .

TABLE 4 Effect of intervention in droplet ranges D – difference between mean value pre and during intervention

Microns	DI	0.3–0.5	0.5–1	1–3	3–5	5–10	> 10	D2	0.3–0.5	0.5–1	1–3	3–5	5–10	> 10
NIV														
Normal	D	-0.097	-0.103	0.096	-0.073	-0.108	0.148	-0.062	-0.065	-0.057	-0.013	0.028	0.100	
	p	0.0991	0.992	0.863	0.653	0.636	0.379	0.968	0.938	0.873	0.544	0.449	0.407	
Patient	D	-0.003	0.004	0.021	0.143	-0.228	0.666	-0.002	-0.002	0.027	0.067	0.165	0.184	
	p	0.565	0.412	0.400	0.231	0.806	0.042	0.470	0.298	0.298	0.308	0.243	0.339	
Coryzal	D	-0.038	-0.068	-0.043	0.175	-0.060	0.807	-0.045	-0.055	0.036	0.233	0.422	0.574	
	p	0.894	0.912	0.639	0.143	0.552	0.044	0.908	0.909	0.238	0.047	0.018	0.052	
Mod NIV														
Normal	D	-0.071	-0.073	-0.054	-0.078	-0.084	-0.086	-0.073	-0.081	-0.089	-0.114	-0.125	0.057	
	p	0.971	0.963	0.703	0.703	0.690	0.588	0.975	0.970	0.963	0.836	0.734	0.431	
Patient	D	-0.020	0.005	0.027	0.013	-0.039	0.087	-0.018	-0.017	-0.003	-0.014	-0.005	0.236	
	p	0.829	0.437	0.393	0.463	0.558	0.402	0.776	0.687	0.521	0.545	0.513	0.244	
Coryzal	D	-0.024	-0.026	-0.011	-0.147	-0.208	0.151	-0.016	-0.037	-0.092	-0.125	-0.153	0.011	
	p	0.776	0.774	0.539	0.861	0.770	0.240	0.671	0.815	0.857	0.813	0.745	0.486	
Oxygen														
Normal	D	-0.063	-0.083	-0.041	-0.003	-0.234	-0.013	-0.050	-0.057	-0.062	-0.049	-0.077	0.081	
	p	0.961	0.942	0.572	0.502	0.576	0.519	0.947	0.902	0.847	0.709	0.713	0.400	
Patient	D	-0.003	-0.009	-0.023	-0.114	-0.068	-0.007	-0.011	-0.027	-0.051	-0.090	-0.102	-0.069	
	p	0.578	0.636	0.577	0.748	0.565	0.507	0.728	0.789	0.736	0.764	0.685	0.554	
Coryzal	D	-0.047	-0.067	-0.075	-0.108	-0.171	-0.012	-0.037	-0.045	-0.032	-0.001	0.004	0.058	
	p	0.955	0.913	0.773	0.750	0.732	0.511	0.905	0.846	0.699	0.503	0.489	0.429	

continued

TABLE 4 Effect of intervention in droplet ranges D – difference between mean value pre and during intervention (continued)

Microns	DI	0.3–0.5	0.5–1	1–3	3–5	5–10	> 10	D2	0.3–0.5	0.5–1	1–3	3–5	5–10	> 10	
Physio															
Patient	D	-0.005	0.057	0.123	0.128	-0.010	1.393		-0.011	0.024	0.070	0.169	0.175	0.424	
	p	0.610	0.118	0.164	0.260	0.511	0.003		0.702	0.206	0.151	0.134	0.228	0.158	
Nebuliser															
Normal	D	15.660	109.480	71.681	27.054	404.932	2.270		25.878	87.932	46.887	1.549	0.232	0.207	
	p	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001		<0.0001	<0.0001	<0.0001	<0.0001	0.120	0.270	
Patient	D	15.516	91.193	56.320	3.967	0.426	0.253		23.080	90.576	45.920	1.642	0.149	0.309	
	p	<0.0001	<0.0001	<0.0001	<0.0001	0.111	0.261		<0.0001	<0.0001	<0.0001	<0.0001	0.241	0.281	
Coryzal	D	11.204	64.822	38.341	1.871	0.197	0.349		17.994	49.458	30.454	1.144	0.234	0.384	
	p	<0.0001	<0.0001	<0.0001	<0.0001	0.229	0.192		<0.0001	<0.0001	<0.0001	<0.0001	0.097	0.133	

DI, distance 1; D2, distance 2; p, p-value.
p-values <0.05 are highlighted in bold text.

Chapter 4

Discussion

The results suggest that NIV using a vented mask in patients with an acute exacerbation of chronic lung disease disseminates large droplets locally. However, at a distance of 1 m the count has fallen significantly. There was no evidence of the generation of small droplets, i.e. an aerosol. Coryzal subjects also produced large droplets that spread for at least 1 m, which indicates that those with rhinorrhoea/upper airway inflammation also generate droplets. This group might be more representative of patients with an early progressive viral infection who are unlikely to produce large volumes of infected sputum when compared with those with cystic fibrosis or bronchiectasis. However, we did not see a difference in counts in those with markedly productive coughs compared with those with minimal sputum production (asthma, obesity hypoventilation) on the day of study. The large droplet count proximal to the mask was significantly reduced in both the patients and coryzal group in the NIV circuit with exhalation port filter, indicating that this modification minimises large droplet dissemination. These filters do not appear to increase the work of breathing if changed regularly. The finding that bilevel NIV with a vented mask disseminates large droplets is in keeping with the superspreading episodes seen in the SARS outbreak, where NIV use was found to be a risk factor on multiple logistic regression analysis.¹²

Physiotherapy has not previously been included in the list of interventions in which PPE and FFP3 masks are indicated. Indeed the results show that it is not an AGP but, perhaps not surprisingly, given that the point of chest physiotherapy is to clear secretions, there was a significant increase in large droplets. As expected, these levels have dropped by D2 but the findings indicate that use of full PPE may be prudent for physiotherapists and respiratory therapists carrying out these procedures in patients with chronic respiratory disease in whom the H1N1 virus has generated an infective exacerbation or secondary bacterial pneumonia.

Oxygen therapy was not associated with an increase in droplets in any group, in any aerosol or droplet range. While it may be possible that 24% O₂ might be an insufficient flow rate to shear droplets from

the upper airway or disseminate those generated by spontaneous coughing or sneezing, we did not see an increase in droplet count in coryzal subjects who used 60% O₂ – a flow rate more typical of that required by patients with acute lung injury due to viral pneumonia. These results should be contrasted with those from Yu *et al.*,¹² who showed O₂ was a significant risk factor for superspreading events. However, their results were based on correlation rather than direct measurement of droplet densities and may be affected by the fact that sicker patients with higher viral loads are more likely to require O₂ than those with milder disease who do not.

The association of spread of SARS with nebuliser use is controversial. Although there are case reports,^{15,16} in this study it is not possible to separate out droplets generated by the nebuliser itself from those generated by the patient. In addition, in clinical practice, patients being treated with nebulised bronchodilator are likely to have air flow obstruction due to asthma or COPD and are therefore more likely to be coughing and wheezing spontaneously. It is plausible that the flow from the nebuliser (either powered by a compressor or oxygen) would disseminate spontaneously generated droplets further. It is notable that the nebuliser was the only intervention that produced in droplets in the aerosol range (< 5 µm). This is entirely in line with the droplet range designed to be generated by this device, and means that this intervention also acts as a quality control confirming that the Aerotrak counters were fully able to detect particles in this range in clinical circumstances. However, in both the coryzal group and the patients we did not detect droplets in the 5- to 10-µm and > 10-µm ranges as occurred during NIV and physiotherapy. This indicates that the vast majority of droplets are likely to be nebulised saline as opposed to patient droplet secretions.

Limitations

We have used droplets as a proxy for viral dissemination, so we do not know whether an increase in droplet count confers an increased risk

of infection for an exposed individual, although we believe this to be biologically plausible. This inference can be confirmed only in viral sampling studies in individuals with influenza, SARS, tuberculosis or other airborne pathogens. This further work would be valuable.

Furthermore, the patients had infective exacerbations of chronic lung disease and the pathology of this is completely different to the acute lung injury that is seen in young patients with normal lungs that are infected with H1N1 or H5N1 influenza. NIV is not indicated in patients with rapidly progressive acute lung injury, although in those with milder disease, and if used earlier in the course of the illness, it might have a role. Emerging evidence suggests that selected cases of H1N1 pneumonia worldwide were treated with NIV with variable results.^{34,35} We believe, however, that the group with chronic lung disease and infectious exacerbations is the most likely to benefit from NIV, and the coryzal group used in this study may reflect airway secretion levels in viral pneumonia patients more closely. However, coryzal patients are clearly less unwell, less dyspnoeic and their lung compliance is likely to be near normal. This is relevant as decreased lung compliance enhanced the dispersion of smoke particles in the human simulator model.³⁶

We sampled droplets at two points – proximal to the subject's nose/mouth/mask and at a 1-m distance. As Hui *et al.*³¹ have shown in smoke particle experiments, flow from mask vents and leaks creates a high to low density vortex, and it is possible that we missed important sampling areas. In order to minimise this risk we used the

information gained from those studies to site D1, the point of maximum density demonstrated by Hui *et al.*,³¹ and placed D2 counter at 1 m, as DH guidelines suggest that health-care workers beyond this distance may use surgical masks as risk of transmission lower. Additionally, we placed D2 at a height of approx 1.52 m (5 ft) equivalent to the nose level of average health-care worker standing 1 m from the patient.

The experiments were carried out in a single room and we minimised disturbances, such as door opening and ventilation, to control the number of variables. Ventilation and air currents are likely to have a significant effect on small size droplets and aerosols, and, indeed, we saw a continued small fall in background count through interventions, which contributes to the mean differences seen in *Table 4*. However, the main impact of treatments (apart from nebuliser) was on large droplets, which, due to greater mass and terminal velocity, will be less affected by air currents.

We have carried out a series of comparisons and have expressed results as mean difference and *p*-values. In the discussion we have used *p*-values of < 0.05 to express significance. It could be argued that adjustments should be made for multiple comparisons. However, we believe the interventions to be independent, and, if comparisons are reduced by either considering one intervention at a time or pooling large versus small droplets, similar conclusions will be reached. We believe, on balance, that it is important to interpret the data erring on the side of caution with respect to risk of dissemination,³⁷ and that these inferences are clinically plausible.

Chapter 5

Conclusions

Despite the limitations, this study indicates that NIV, O₂ and physiotherapy are not AGPs. Physiotherapy and NIV generate large droplets adjacent to the patient, but these fall significantly at 1 m from the patient. A mod NIV circuit using a non-vented mask and filtered exhalate reduces the number of large droplets produced. Nebulised saline delivered by a mouthpiece produces an aerosol of droplets, but most are in the expected droplet range for the device and large droplets were not seen in patients and coryzal subjects. O₂ at 60% and 24% did not appear to be an aerosol or droplet-generating procedure.

What are the implications for clinical practice and infection control? These results imply that during NIV and physiotherapy use of full PPE should be considered for health-care team members working within 1 m of the patient, as droplet count is increased. As the droplets are large and many drop out within 1 m over bedside surfaces, the crucial importance of handwashing and decontamination of near surfaces is evident, as transmission in these

circumstances is likely to be direct via droplet spread or from fomites and direct contact with the patient's local environment. As small and aerosolised particles were not demonstrated, the role other protective measures, such as negative pressure rooms, which have been advocated in some pandemic flu guidelines, may be less important.

Recommendations for further research:

1. Droplet sampling should be carried out in patients with pandemic influenza to confirm that droplets generated in this situation are comparable to those produced by patients in this study.
2. Droplet sampling sizing could be carried out in human simulator models with laser droplet imaging to corroborate results.
3. Viral carriage in different size droplets should be assessed to test whether using droplets as a proxy of infectivity risk is a realistic clinical substitute.



Acknowledgements

With thanks also to Winston Banya, Senior Statistician, Research Department, and Professor Paul Cullinan, Occupational Medicine Department, Royal Brompton Hospital, who provided statistical advice. We are grateful to the NIHR HTA panel for suggesting the addition of the coryzal group to study design.

The project was supported by the NIHR Respiratory Disease Biomedical Research Unit at the Royal Brompton and Harefield NHS Foundation Trust and Imperial College London.

Contribution of authors

AK Simonds, Consultant and Reader in Respiratory Medicine, Royal Brompton & Harefield NHS Foundation Trust, designed the study, wrote the grant application, analysed results with KP, JS and RD, and wrote the report.

A Hanak, Senior Respiratory Support Technician and Physiotherapist, Royal Brompton & Harefield NHS Foundation Trust, carried out the experiments, including performing physiotherapy, and created and maintained the database.

M Chatwin, Consultant Physiotherapist, Royal Brompton & Harefield NHS Foundation Trust, carried out the experiments, including performing

physiotherapy, performed pilot studies and recruited patients.

MJ Morrell, Reader in Sleep Physiology, National Heart & Lung Institute, Imperial College, contributed to study design and critical analysis of results.

A Hall, Consultant in Microbiology & Infection Control Lead, Royal Brompton & Harefield NHS Foundation Trust, contributed to study design, provided infection control and microbiology advice, and critical analysis of results.

KH Parker, Professor Physiological Fluid Mechanics (Emeritus) Department of Bioengineering, Imperial College, provided guidance on droplet characterisation and mathematical analysis, analysed results, provided statistical advice and critical review, and contributed to writing the report and figures.

JH Siggers, Lecturer, Department Bioengineering, Imperial College, advised on study design, mathematics of droplet analysis, critical review of results and contributed to writing the report.

RJ Dickinson, Senior Lecturer, Department of Bioengineering, Imperial College, advised on study design, droplet analysis, critical review of results and contributed to writing the report.



References

1. Brankston G, Gitterman L, Hiriji Z, Lemieux C, Gardam M. Transmissin of influenza A in human beings. *Lancet Infect Dis* 2007;**7**:257–65.
2. Tellier R. Review of aerosol transmission of influenza A virus. *Emerg Infect Dis* 2006;**12**:1657–62.
3. Fabian P, McDevitt JJ, DeHaan WH, Fung ROP, Cowling BJ, Chan KH, *et al.* Influenza virus in human exhaled breath: an observational study. *PLoS ONE* 2008;**3**:e2691.
4. The ANZIC Influenza Investigators. Critical care services and 2009 H1N1 influenza in Australia and New Zealand. *N Engl J Med* 2009;**361**:1925–34.
5. Jain S, Kamimoto L, Bramley AM, Schmitz AM, Benoit SR, Louie J, *et al.* Hospitalised patients with 2009 H1N1 influenza in the United States, April–June 2009. *N Engl J Med* 2009;**361**:1935–44.
6. Department of Health. *Pandemic flu: a national framework for responding to an influenza pandemic*. London: Department of Health; 2009.
7. Department of Health. *Pandemic influenza: managing demand and capacity in healthcare organisations (surge)*. London: Department of Health; 2009.
8. Department of Health and Health Protection Agency. *Pandemic (H1N1) 2009 influenza. A summary of guidance for infection control in healthcare settings*. London: Department of Health; 2009.
9. Fowler RA, Guest CB, Lapinsky SE, Sibbald WJ, Louie M, Tang P, *et al.* Transmission of severe acute respiratory syndrome during intubation and mechanical ventilation. *Am J Respir Crit Care Med* 2004;**169**:1198–202.
10. Scales DC, Green K, Chan AK, Poutanen SM, Foster D, Nowak K, *et al.* Illness in intensive care staff after brief exposure to severe acute respiratory syndrome. *Emerg Infect Dis* 2003;**9**:1205–10.
11. Lau JTF, Fung KS, Wong TW, Kim JH, Wong E, Chung S, *et al.* SARS transmission among hospital workers in Hong Kong. *Emerg Infect Dis* 2004;**10**:280–6.
12. Yu IT, Xie Z, Tsoi K. Why did outbreaks of severe acute respiratory syndrome occur in some hospital wards and not others? *Clin Infect Dis* 2007;**44**:1017–25.
13. Sokol DK. Virulent epidemics and scope of healthcare workers' duty of care. *Emerg Infect Dis* 2006;**12**:1238–41.
14. Simonds AK, Sokol DK. Lives on the line? Ethics and practicalities of duty of care in pandemics and disasters. *Eur Respir J* 2009;1–7.
15. Lee N, Hui DS, Wu A, Chan P, Cameron P, Joynt GM, *et al.* A major outbreak of severe acute respiratory syndrome in Hong Kong. *N Engl J Med* 2003;**348**:1986–94.
16. Wong RS, Hui DS. Index patient and SARS outbreak in Hong Kong. *Emerg Infect Dis* 2004;**10**:339–41.
17. Simonds AK. Continuous positive airway pressure and non-invasive ventilation in acute hypoxaemic respiratory failure. In Simonds AK, editor. *Non-invasive respiratory support: a practical handbook*. London: Hodder Arnold; 2007. pp. 57–71.
18. McCracken J. Should noninvasive ventilation be considered a high-risk procedure during an epidemic? *CMAJ* 2009;**181**:663–4.
19. World Health Organization (WHO). *Clinical management of human infection with influenza A (H5N1) virus*. Geneva: WHO; 2007. Advice updated 15 August 2007.
20. Gomersall CD, Tai DYHT, Derrick JL, Goh MS, Buckley TA, Chua C, *et al.* Expanding ICU facilities in an epidemic: recommendations based on experience from SARS epidemic in Hong Kong and Singapore. *Intensive Care Med* 2006;**32**:1004–13.
21. World Health Organization (WHO) *Interim Guidelines. Infection prevention and control of epidemic- and pandemic-prone acute respiratory diseases in health care*. Geneva: WHO; 2007.
22. Cheung TMT, Yam LYC, So LKY, Lau ACW, Poon E, Kong BMH, *et al.* Effectiveness of noninvasive positive pressure ventilation in the treatment of acute respiratory failure in severe acute respiratory syndrome. *Chest* 2004;**126**:845–50.
23. Yam LYC, Chan AYF, Cheung TMT, Tsui ELH, Chan JCK, Wong VCW. Non-invasive versus invasive mechanical ventilation for respiratory failure in severe acute respiratory syndrome. *Chin Med J* 2005;**118**:1413–21.

24. Department of Health. *Pandemic influenza: surge capacity and prioritisation in health services*. London: Department of Health; 2008.
25. Confalonieri M, Potena A, Carbone G, Della Porta R, Tolley E, Meduri GU. Acute respiratory failure in patients with severe community-acquired pneumonia. A prospective randomised evaluation of noninvasive ventilation. *Am J Respir Crit Care Med* 1999;**160**:1585–91.
26. Seto WH, Tsang D, Yung RW, Ching TY, Ng TK, Ho M, *et al*. Effectiveness of precautions against droplets and contact in prevention of nosocomial transmission of severe acute respiratory syndrome (SARS). *Lancet* 2003;**361**:1519–20.
27. Somogyi R, Vesely AE, Azami T, Preiss D, Fisher J, Fowler RA. Dispersal of respiratory droplets with open vs closed oxygen delivery masks: implications for transmission of severe acute respiratory syndrome. *Chest* 2004;**125**:1157.
28. Hui DS, Hall SD, Chan MTV, Chow BK, Tsou JY, Joynt GM, *et al*. Noninvasive positive-pressure ventilation: an experimental model to assess air and particle dispersion. *Chest* 2006;**130**:730–40.
29. Ip M, Tang JW, Hui DS, Wong ALN, Chan MTV, Joynt GM, *et al*. Airflow and droplet spreading around oxygen masks: a simulation model for infection control research. *Am J Infect Control* 2007;**35**:684–9.
30. Mardimae A, Slessarev M, Han J, Sasano H, Sasano N, Azami T, *et al*. Modified N95 mask delivers high inspired oxygen concentrations while effectively filtering aerosolized microparticles. *Ann Emerg Med* 2006;**48**:391–9.
31. Hui DS, Chow BK, Ng SS, Chu LCY, Hall SD, Gin T, *et al*. Exhaled air dispersion distances during non-invasive ventilation via different Respironics facemasks. *Chest* 2009;**136**:998–1005.
32. Hui DS, Hall SD, Chan MTV, Chow BK, Ng SS, Gin T, *et al*. Exhaled air dispersion during oxygen delivery via a simple oxygen mask. *Chest* 2007;**132**:826.
33. Tang JW, Liebner TJ, Craven BA, Settles GS. A Schlieren optical study of the human cough with and without wearing masks for aerosol infection control. *J R Soc* 2009;**S727–36**.
34. Rello J, Rodriguez A, Ibanez P, Socias L, Cebrian J, Marques A, *et al*. Intensive care of adult patients with severe respiratory failure caused by influenza A (H1N1) in Spain. *Critical Care* 2009;**13**:R148.
35. Kumar A, Zarychanski R, Pinto R, Cook DJ, Marshall J, Lacroix J. Critically ill patients with 2009 influenza A(H1N1) infection in Canada. *JAMA* 2010;**302**:1872–9.
36. Hui DS, Chow BK, Chu LCY, Ng SS, Hall SD, Gin T, *et al*. Exhaled air and aerosolized droplet dispersion during application of a jet nebuliser. *Chest* 2009;**135**:648–54.
37. Perneger TV. What's wrong with Bonferroni adjustments. *BMJ* 1998;**316**:1236–8.

Appendix I

Protocol

Evaluation of droplet dispersion during NIV, O₂ and nebulised drug delivery in clinical practice

- Project management members:
 - *Chief investigator* Dr Anita K Simonds, Academic Department Sleep & Breathing, Royal Brompton Hospital, Sydney Street, London SW3 6NP, 020 73518911
 - *Co-investigators* Dr R Dickinson and Dr J Siggers, Bioengineering Department, IC, Dr Michelle Chatwin & Dr Anne Hall RBH, Dr M Morrell, IC
 - *Statistician* Mr Winston Banya
 - *Project manager* Dr Michelle Chatwin, 020 73518911
 - *Key contact* Dr Anita K Simonds (A.Simonds@rbht.nhs.uk)
- Funder: NIHR HTA
- Sponsor: Royal Brompton & Harefield NHS Foundation Trust
- Contact: Wendy Butcher, R&D Department, Royal Brompton Hospital

Introduction

Background

Influenza viruses are spread by droplets, but aerosols may be implicated. While many patients recover without serious illness, some with H1N1 swine flu will develop pneumonia/respiratory insufficiency requiring treatment by oxygen therapy (O₂), ventilatory support or nebulised drugs, and this is more likely in those with underlying respiratory or cardiac disorders, for example chronic obstructive pulmonary disease (COPD), asthma, cystic fibrosis, genetic susceptibility, pregnancy or if the virus mutates. These therapies may generate droplets or aerosol during delivery, which, in the severe acute respiratory syndrome (SARS) outbreak, were associated with an increased incidence of infection in health-care workers (Fowler *et al.* 2004)⁹ and superspreading events on hospital wards (Yu *et al.* 2007).²⁴ Non-invasive ventilation (NIV) is unlikely to be effective in patients with overwhelming acute lung injury, but in early pneumonia and in those in whom influenza has caused an exacerbation of COPD or heart failure NIV may be effective in reducing the need for intensive care unit (ICU)

admission. Currently, use of NIV in pandemic flu is controversial. Department of Health Pandemic Influenza guidance recommends that NIV should be used with full infection control (aerosol-generating) precautions by experienced units employing practice guidelines that have been developed by our team (Simonds 2007),²⁵ but there is no substantive evidence base and NIV use is not advocated in other national guidelines. Hui *et al.* (2006) carried out studies of NIV droplet distribution using a patient simulator and smoke particles, but there have been no systematic studies in humans or during oxygen and nebuliser therapy, or physiotherapy.

Rationale for current study

This research should provide the first analysis of droplet distribution around respiratory therapies in clinical circumstances that are relevant to H1N1 infection. Although the patients with chronic respiratory disease will not specifically have an exacerbation triggered by H1N1 influenza in this study, the results should be representative of any acute exacerbation and we will also study those with coryzal symptoms, some of whom may have H1N1 infection. The findings should enable health-care professionals to understand patterns of geographical distribution of respirable droplets when caring for patients, inform selection of circuitry and interfaces to reduce dissemination, and by modelling the profile of decay of particles after therapy we hope to guide health-care workers' entry into rooms of unstable patients.

Impact on practice

As this is the first analysis of distribution of droplets during NIV, O₂ and nebuliser therapy in representative clinical circumstances, the results obtained should influence clinical practice and policy immediately by:

1. informing the choice of interface/delivery systems
2. guiding health-care workers to safer application in pandemic flu and enable them to understand relative risks
3. reducing the risk of dissemination to other patients and staff in superspreading events

4. wider, safe use of NIV may reduce ICU bed pressures, as NIV may be performed in respiratory ward areas/high-dependency single rooms.

Study objectives

The key objective is to understand the characteristics of droplet and aerosol dispersion around delivery systems during NIV, O₂ therapy, nebuliser therapy and physiotherapy procedures.

We will examine:

1. droplet size and count
2. geographical distribution of droplets
3. rise and decay of droplets over time after the therapies are initiated and discontinued
4. the impact of modifications to the delivery system to reduce droplet/aerosol dissemination in:
 - i. normal subjects
 - ii. individuals with coryzal symptoms
 - iii. patients with an acute exacerbation of chronic lung disease.

Primary objective

- To evaluate the characteristics of droplets and aerosol generated using NIV, O₂ therapy, nebuliser therapy and physiotherapy in clinical practice.

Secondary objectives

- To determine whether particular delivery methods/interfaces generate more droplets.
- To establish how can droplet characteristic information be applied to inform safe use of these therapies in patients with H1N1 swine flu, and other droplet/aerosol-borne diseases.

Study methodology

Overall design

This is an observational study with subjects and patients acting as their own control.

Setting and timescale

The study will be carried out in a single centre (Royal Brompton Hospital) over 4 months (September to December 2009).

Study outcome measures

Number of droplets in size range 0.3–10 µm, measured during conditions listed below.

Specific methods

Droplet count and sizing

We will count droplets in size range 0.3–10.0 µm within distributions of 0.3–0.5, 0.5–1.0, 1.0–3.0, 3.0–5.0, 5.0–10.0 and > 10 µm using Aerotrak Model 8220 optical particle counter with counting efficiency 50% ± 10% at 0.3 µm and 100% ± 10% at 0.45 µm and greater. We will examine dissemination of smaller droplet (aerosol) size using a P-Trak Ultrafine Condensation particle counter (particle size range 0.02–1.0 µm) at sample flow rate 100 cm³/minute. Each sampling will be carried out twice over 10 seconds, on three occasions, at sampling points: (1) adjacent (within 2 cm) to mouth or mask; (2) 0.5 m from mouth or mask; (3) 1 m from mouth or mask; and (4) 3 m from mouth or mask with sampling points (2)–(4) being carried out in radial positions – two laterally to subject/patient, one directly in front of subject/patient and one above subject. The Aerotrak and P-Trak counter devices will be mounted on tripods to maintain accuracy and reproducibility of measurements.

Mathematical modelling

We will use mathematical modelling of droplet motion and dispersion to derive the expected droplet distribution at different distances. Fitting the model with observations at a number of positions will allow interpolation and extrapolation of the measured droplet distribution as a function of size of the droplet and distance from the patient–mask interface, for a range of room conditions. In turn, this will enable us to predict the safe times and distances beyond which exposure can be considered comparable to spontaneous breathing or negligible.

Participants

Groups

We will study three groups: normal subjects, subjects with coryzal (common cold or flu-like) symptoms and adult patients with chronic lung disorders.

Inclusion criteria

Normal subjects Age 18 years and above. Able to speak English and understand protocol.

Coryzal subjects Age 18 years and above. Have two of any of following: raised temperature or history of raised temperature, sore throat, headache, muscle aches and pains, cough in previous

24–48 hours. Arterial oxygen saturation 95% or above on air.

Patients A clinical diagnosis confirmed by medical consultant of COPD, asthma, cystic fibrosis, bronchiectasis, chest wall disorder or neuromuscular disease, for example Duchenne muscular dystrophy. Admitted with infective exacerbation defined by increased breathlessness, raised white cell count or temperature or CRP (C-reactive protein – raised values indicate infection or inflammation). Requiring treatment with O₂ and NIV as clinically indicated.

Exclusion criteria

Normal subjects Current illness or underlying chronic condition. Pneumothorax in previous 3 months. Unable to understand English or trial information.

Coryzal subjects Underlying chronic condition. Arterial oxygen saturation < 95% on air. Pneumothorax in previous 3 months. Unable to understand English or trial information.

Patients Haemodynamically unstable (systolic blood pressure < 90 mmHg, uncontrolled arrhythmia), medically unstable, arterial oxygen tension (PaO₂) < 7.5 kPa on O₂ or NIV, arterial carbon dioxide tension (PaCO₂) > 7.5 kPa on NIV or O₂, unable to breathe spontaneously for < 4 hours. Unable to understand English or trial information.

Sampling method

Normal subjects Will be recruited from departmental database of normal subjects who have participated in previous studies.

Coryzal subjects Will be recruited from occupational health department, and staff who develop symptoms while on duty.

Patients Will be recruited from those already inpatients on respiratory ward, with an acute infective exacerbation of chronic lung disease. At any one time we have around 15–20 patients on the ward receiving O₂/NIV. The research team are either members of the clinical team or they interact with the team on a daily basis.

Sample size

Background:

- It should be stressed that this work is almost exclusively exploratory in nature. This is because there are very many unknowns.

- It is not known whether the material generated by infected individuals breathing, coughing or undergoing interventions is in the form of a fine aerosol or larger droplets.¹ NIV and nebulisation have been termed ‘potential aerosol-generating procedures’ but this is based on presumption, not evidence. In the Department of Health Pandemic Flu guidelines³ it is stipulated that high-efficiency masks should be used when working within 1 m of the patient, and that beds of patients being cohort nursed should be more than 1 m apart. There is little primary evidence for either of these stipulations but in the SARS outbreak superspreading events (i.e. at least three cases arising from one index case) were associated with a distance between beds of < 1 m and index cases with the use of O₂ or non-invasive ventilation.²⁴
- Further, the ‘dose’ needed to infect is not clear as droplets are a proxy measure of virus presence/infectivity, and sicker patients with higher viral loads are likely to need more therapeutic interventions.
- Moreover we do not know the rate of decay of droplets over time after interventions have been discontinued. Again, this will be partly related to size as larger droplets with greater mass will more quickly fall to the floor or onto bedding.

Droplet size and number – pilot data, variability and clinically meaningful difference:

- We have pilot data from five normal subjects sampled at the mouth or mask and in one droplet size range (5–10 µm). This size range is known as the ‘respirable range’, representing droplets likely to be deposited in lungs; larger droplets are not inspired and very small aerosol particles do not have sufficient mass to drop out in lung and are expired as easily as they are inspired. Droplets generated by interventions (O₂, NIV, etc.) should be compared with those generated by the subject/patients breathing spontaneously, as a baseline of zero droplets is not clinically realistic. We are therefore carrying out comparisons with spontaneous breathing with each subject/patient acting as their own control.
- Our pilot data above estimated a droplet count of 900 (standard deviation = 100) with spontaneous breathing.
- In the absence of any other published information and the uncertainties outlined above, we have chosen a doubling in this

droplet count to represent a significant increase in risk of spread to health-care staff or other patients. This estimate is informed by the observation that coughing and sneezing in pilot work resulted in a count of around 1800, and that coughing and sneezing increase the risk of infection.

We used Statistical Analysis Systems (SAS), version 9.1, to estimate the required study group sizes:

- Using our pilot estimates and a false-positive rate (α) of 0.05, calculations for a single two-group comparison with 80% power indicate that very small groups would be required.
- We have, however, increased our group sizes to account for the possibility that variability may be higher in patients (currently unknown) and the four comparisons that are to be undertaken. Sample sizes are therefore normal subjects = 10, coryzal subjects = 10, patients = 20.
- This model is based on droplet counts in one size range at the mouth and is suitable for our primary purpose. Again, with the lack of any information from elsewhere, we do not know whether our sample size will be sufficient for our other questions: for example, number of droplets at different distances from patient or the decay over time. Initial findings will provide further information. If variability estimates are greater or differences smaller compared to spontaneous breathing, further recruitment will be possible.

Statistical advice was provided by Mr Winston Banya, Senior Statistician, R&D Department, Royal Brompton Hospital.

Preregistration evaluations

We will check that arterial oxygen saturation level is 95% or above in normal subjects and subjects with coryzal symptoms using an oximeter ear probe. In coryzal subjects nasopharyngeal aspiration will be carried out along with throat and nasal swabs. Virology results will not be known until after study tests are done, so they will inform the analysis but are not needed for study entry as symptoms alone determine eligibility.

Patients will have an arterial oxygen saturation value of more than 88% and TcCO₂ value of less than 7.5 kPa on O₂ and or NIV.

Withdrawal criteria

The trial will be discontinued if the chief investigator feels it is unsafe to continue. As the therapies used are in routine clinical practice in patients, and researchers are members of the clinical team and routinely apply these therapies in patients, including those in first wave of swine flu, this risk is relatively low.

Recruitment and methodological process

Recruitment

Recruitment will take part at the Royal Brompton Hospital. Normal subjects will be recruited from departmental database and volunteers working in the hospital. Coryzal subjects will be recruited from the occupational health department and from individuals working in the hospital who develop symptoms while on duty.

Written informed consent will be obtained by the research fellow, Dr Michelle Chatwin or CI at the Royal Brompton Hospital, who have all had training in obtaining consent. Subjects and patients will be provided with information sheets. Normal subjects and patients will have 24 hours to decide whether to participate and coryzal subjects will have 1 hour to decide.

Methodological process

- This is an observational trial that will be carried out in a single hospital side room on respiratory ward at the Royal Brompton Hospital. The aim is to measure the size and number of droplets and smaller (aerosol) particles generated during treatment with NIV, O₂, nebuliser therapy and during physiotherapy.

Three groups will take part:

- (A) normal subjects
- (B) subjects with coryzal (common cold or flu-like) symptoms
- (C) patients with respiratory insufficiency due to COPD, cystic fibrosis, chronic asthma, bronchiectasis, neuromuscular disease receiving NIV/O₂/nebuliser therapy as indicated for an infective exacerbation. Each subject or patient will take part on one occasion, the study taking approximately 3 hours to complete.

Subjects and patients:

- (A) Normal subjects will be recruited from our database of normals (aged 18 years and above) and above. *Exclusion criteria:* no current illness or underlying chronic condition.
- (B) Individuals with common cold or flu-like (coryzal) symptoms defined by pyrexia, and two of sore throat, muscle aches and pains, headache, cough within previous 24–48 hours (age 18 years and above) will be recruited from contacts from normal patient database, occupational health department of the Royal Brompton Hospital and from staff developing symptoms while on duty. They will be studied after having nasopharyngeal swabs for viral screening, to confirm diagnosis. *Exclusion criteria:* no underlying chronic health conditions, medically stable.
- (C) Patients with chronic respiratory failure will be recruited from those admitted to the ward with an infective exacerbation of chronic respiratory disease. Inclusion criteria: those with COPD, cystic fibrosis, bronchiectasis, chest wall disorder and neuromuscular disease. These groups are selected as will contain older patients with COPD and younger patients with cystic fibrosis and, for example, Duchenne muscular dystrophy, in whom NIV and O₂ therapy is clinically indicated. *Exclusion criteria:* haemodynamically or medically unstable, PaO₂ < 7.5 kPa, PaCO₂ > 7.5 kPa pH < 7.34 on therapy, cognitive inability to able to understand study information sheet, able to breathe spontaneously for < 4 hours.

Technologies being assessed:

- Non-invasive ventilation using standard bilevel pressure support device with a range of interfaces and settings, nasal continuous positive airway pressure (CPAP) therapy, O₂ therapy via 60%, 35% and 24% masks

Measurements:

- Droplets will be visualised using a Model 8220 Aerotrak Optical particle counter (TSI Inc.) with particle size detection of 0.3–10 µm, and a Model 8525 P-Trak Ultrafine Condensation particle counter (TSI Inc.) adjacent to subject/delivery system, 1 m from delivery system and 3 m from patient/subject, at six fixed radial points.

Investigation plan:

- On arrival in the side room, subjects and patients will be assessed breathing spontaneously at rest, during simulated coughing, and then, when receiving NIV and O₂, physiotherapy and nebulised saline therapy in random order.
- Droplet distribution will be measured in the following test conditions (selected as clinically representative).
 1. For (A) normal subjects and (B) subjects with coryzal symptoms:
 - Control* Spontaneous breathing and simulated cough with and without surgical mask, which will take approximately 10 minutes.
 - Non-invasive ventilation* A bilevel ventilator will be used: in random order delivery with non-vented full-face mask, total face mask and helmet with and without filter modification and vented full-face mask. Ventilator settings: inspiratory positive airway pressure (IPAP), expiratory airway pressure (EPAP), IPAP/EPAP 20/5 15/5 10/5 cmH₂O. CPAP 5 and 10 cmH₂O. This will take approximately 1 hour.
 - O₂ therapy* Will be delivered using 60%, 35%, 24% masks. This will take approximately 30 minutes. This will take about 20 minutes.
 - Nebulised 0.9% saline* Delivered from standard nebuliser. This will take 10 minutes.
 - Standardised physiotherapy* This will take 10 minutes.

Subjects will be able to have rest periods between the runs, as we will be sampling the room to ensure control conditions obtain and get background counts.

2. For (C) patients with respiratory insufficiency:
 - Spontaneous breathing and during simulated cough* This will take approximately 10 minutes.
 - Non-invasive ventilation* Using current clinically indicated NIV settings delivered in random order through non-vented full-face mask, total face mask, helmet with and without filter modification and vented mask. This will take approximately 45 minutes.

- iii. *O₂ therapy 24% Ventimask* spontaneously breathing. This will take approximately 5–10 minutes.
- iv. *Nebulised 0.9% saline* Delivered by standard nebuliser. This will take approximately 10 minutes.
- v. *During physiotherapy using 24% O₂ mask* This will take about 10 minutes.

Patients will be monitored with arterial oxygen saturation (SaO₂), transcutaneous carbon dioxide tension (TcCO₂) and heart rate measurement using a non-invasive ear probe (Tosca) throughout stages (i)–(iv).

They will be able to have rest periods between the runs as we will be sampling the room to ensure control condition obtain and get background counts.

Droplet and aerosol characterisation:

- We will count droplets in size range 0.3–10.0 μm within distributions 0.3–0.5, 0.5–1.0, 1.0–3.0, 3.0–5.0, 5.0–10.0 and > 10 μm using Aerotrak Model 8220 optical particle counter with counting efficiency 50% ± 10% at 0.3 μm and 100% ± 10% at 0.45 μm and greater. We will examine dissemination of smaller droplet (aerosol) size using a P-Trak Ultrafine Condensation particle counter (particle size range 0.02–1.0 μm) at sample flow rate 100 cm³/minute. Each sampling will be carried out twice over 10 seconds, on three occasions, at sampling points: (1) adjacent (within 2 cm) to mouth or mask; (2) 0.5 m from mouth or mask; (3) 1 m from mouth or mask; and (4) 3 m from mouth or mask, with sampling points (2)–(4) being carried out in radial positions – two laterally to subject/patient, one directly in front of subject/patient and one above subject. The Aerotrak and P-Trak counter devices will be mounted on tripods to maintain accuracy and reproducibility of measurements.

Mathematical modelling:

- We will use mathematical modelling of droplet motion and dispersion to derive the expected droplet distribution at different distances. Fitting the model with observations at a number of positions will allow interpolation and extrapolation of the measured droplet distribution as a function of size of the droplet and distance from the patient–mask interface,

for a range of room conditions. In turn, this will enable us to predict the safe times and distances beyond which exposure can be considered comparable to spontaneous breathing or negligible.

Equipment:

- Non-invasive ventilation: we will use a Saime Elisee bilevel ventilator, which can deliver a variety of IPAP and EPAP and fixed-level CPAP through a single-limb circuit and a double-limb circuit. The pressures of IPAP/EPAP 20/5, 15/5 and 10/5 cmH₂O (spontaneous triggered mode) and CPAP 5 and 10 cmH₂O have been selected as clinically representative. These pressures will be used in normal subjects and those with coryzal symptoms. In the patient group we will use the IPAP/EPAP settings and back-up respiratory rate as clinically indicated.

Interfaces:

- We will use a full-face masks (ResMed), non-vented with filtered (intersurgical) exhalation port, and vented masks (ResMed), and total masks (Respironics/Philips) in all subjects and patients, and, in five subjects, a helmet (Rusch).

Oxygen therapy:

- Oxygen therapy 60% and 35% via high-flow reservoir mask, 24% via Venturi mask in normal subjects and those with coryzal symptoms, 24% via Venturi mask in patients.

Physiotherapy:

- Will be standardised as cycles of deep breathing, with percussion or shaking to loosen any secretions, followed by an assisted cough initiated manually, augmented by a physiotherapist performing inwards and upwards pressure on the lower thorax to aid expectoration, after which the patient rests and the cycle repeated as required. It will be performed by one physiotherapist (MC) who has performed standardised physiotherapy manoeuvres in other randomised crossover trials.

Nebuliser:

- Actineb nebuliser (Clement Clark) generating droplets of 3–10 μm of 0.9% saline.

There will be an interim analysis and mathematical modelling after 10 subjects and 10 patients have been studied.

Non-invasive ventilation, O₂, nebuliser therapy and standardised physiotherapy will be delivered by research fellow and Dr Michelle Chatwin.

Ethical considerations

The main risk is to research staff in the dissemination of H1N1 and other coryzal viruses. Full personal protective equipment will be used – the research team members are fully familiar with this and have experience in managing H1N1 patients. Some team members have already had swine flu themselves so will be immune.

There is a very small risk of a subject or patient using NIV developing a pneumothorax. The patients already will be using NIV as part of their clinical management.

Adverse events

Potential adverse events:

- Research team member becoming infected with swine flu. The individual would be withdrawn from doing the project and treated with oseltamivir in the normal way. In practice it will be difficult to establish if the individual was infected during the study, by contact with other infected patients or from contact from within or outside the hospital

All adverse events will be reported. Depending on the nature of the event the reporting procedures below will be followed. Any questions concerning adverse event reporting will be directed to the chief investigator in the first instance.

Non-serious adverse events

All such events, whether expected or not, will be recorded.

Serious adverse events

An SAE form should be completed and faxed to the chief investigator within 24 hours. However, hospitalisations for elective treatment of a pre-existing condition will not be reported as SAEs.

All SAEs will be reported to the REC overseeing the research and the research sponsor, where, in the opinion of the chief investigator, the event was:

- *'related'* i.e. resulted from the administration of any of the research procedures, or
- *'unexpected'* i.e. an event that is not listed in the protocol as an expected occurrence.

Reports of related and unexpected SAEs will be submitted within 15 days of the chief investigator becoming aware of the event, using the COREC SAE form for non-IMP studies.

Investigators will report any SAEs as required by their local research ethics committee and/or research and development office.

Assessment and follow-up

We do not plan to follow up patients after the study. Virology results will be fed back to coryzal subjects and appropriate action advised.

Statistics and data analysis

Data will be analysed using ANOVA with correction for repeated measure. Statistical advice will be provided by Mr Winston Banya, R&D Department, Royal Brompton & Harefield NHS Foundation Trust.

Data and all appropriate documentation will be stored for a minimum of 5 years after the completion of the study, including the follow-up period.

Regulatory issues

Ethics approval

The chief investigator will obtain ethical approval from a research ethics committee via the IRAS system. The study will not commence until ethical approval is obtained, and will be conducted in accordance with the recommendations for physicians involved in research on human subjects adopted by the 18th World Medical Assembly, Helsinki 1964, and later revisions.

Consent

Consent to enter the study will be sought from each participant only after a full explanation has been given, an information leaflet offered and time allowed for consideration. Signed participant consent will be obtained. The right of the participant to refuse to participate without giving reasons will be respected. All participants are free to withdraw at any time from the research without giving reasons and without prejudicing further treatment. Consent will be obtained by the patient's existing clinical consultant.

Confidentiality

The chief investigator will preserve the confidentiality of participants taking part in the study in line with the Data Protection Act 1998.

Indemnity

NHS indemnity cover.

Sponsor

Royal Brompton & Harefield NHS Foundation Trust.

Funding and costs

NIHR HTA will fund this study. Travel costs to £20 are available to normal and coryzal subjects.

Audits and inspections

Sponsor and other regulatory bodies will ensure adherence to Good Clinical Practice and the NHS *Research Governance Framework for Health and Social Care* (2nd edn).

Study management

The day-to-day management of the study will be co-ordinated through by Dr Michelle Chatwin (M.Chatwin@rbht.nhs.uk).

Publication policy

Results from this research will be reported and disseminated via peer-reviewed journals and via conference presentations. No personal or identifiable data will be present in any public reports of this research.

References

1. Tellier R. Review of aerosol transmission of influenza A virus. *Emerg Infect Dis* 2006;**12**:1657–62.
2. Fabian P, McDevitt JJ, DeHaan WH, Fung ROP, Cowling BJ, Chan KH, *et al.* Influenza virus in human exhaled breath: an observational study. *PLoS ONE* 2008;**3**:e2691.
3. Department of Health. *Pandemic influenza: guidance for infection control in critical care*. London: Department of Health; 2008.
4. Simonds AK, Sokol DK. Lives on the line? Ethics and practicalities of duty of care in pandemics and disasters. *Eur Respir J* 2009;**34**:303–9.
5. Department of Health. *Pandemic flu. Managing demand and capacity in health care organisations (surge)*. London: Department of Health; 2009.
6. Gomersall CD, Tai DYHT, Derrick JL, Goh MS, Buckley TA, Chua C, *et al.* Expanding ICU facilities in an epidemic: recommendations based on experience from SARS epidemic in Hong Kong and Singapore. *Intensive Care Med* 2006;**32**:1004–13.
7. Guidelines from the British Infection Society, British Thoracic Society and DH. Pandemic flu: clinical management of patients with influenza-like illness during an influenza pandemic. *Thorax* 2007;**62**:1–46.
8. Levy M, Tanios MA, Nelson D, Short K, Senechia A, Vespia J, *et al.* Outcomes of patients with do-not-intubate orders treated with noninvasive ventilation. *Crit Care Med* 2004;**32**:2002–7.
9. Fowler RA, Guest CB, Lapinsky SE, Sibbald WJ, Louie M, Tang P, *et al.* Transmission of severe acute respiratory syndrome during intubation and mechanical ventilation. *Am J Respir Crit Care Med* 2004;**169**:1198–202.
10. Scales DC, Green K, Chan AK, Poutanen SM, Foster D, Nowak K, *et al.* Illness in intensive care staff after brief exposure to severe acute respiratory syndrome. *Emerg Infect Dis* 2003;**9**:1205–10.
11. Xiao Z, Li Y, Chen RC, Li SY, Zhong SQ, Zhong NS, *et al.* A retrospective study of 78 patients with severe acute respiratory syndrome. *Chin Med J* 2003;**116**:805–10.
12. Varia M, Wilson S, Sarwal S, McGeer A, Gournier E, Galanis E, *et al.* Investigation of a nosocomial outbreak of severe acquired respiratory syndrome (SARS) in Toronto, Canada. *CMAJ* 2003;**169**:285–92.
13. Cheung TMT, Yam LYC, So LKY, Lau ACW, Poon E, Kong BMH, *et al.* Effectiveness of noninvasive positive pressure ventilation in the treatment of acute respiratory failure in severe acute respiratory syndrome. *Chest* 2004;**126**:845–50.
14. Yam LYC, Chan AYE, Cheung TMT, Tsui ELH, Chan JCK, Wong VCW. Non-invasive versus invasive mechanical ventilation for respiratory failure in severe acute respiratory syndrome. *Chin Med J* 2005;**118**:1413–21.
15. Yu IT, Li Y, Wong TW, Tam W, Chan AT, Lee JH, *et al.* Evidence of airborne transmission of severe acute respiratory syndrome virus. *N Engl J Med* 2004;**350**:1731–9.
16. Hugonnet S, Pittet D. Transmission of severe acute respiratory syndrome in critical care: do we need a change? *Am J Respir Crit Care Med* 2004;**169**:1177–8.
17. Seto WH, Tsang D, Yung RW, Ching TY, Ng TK, Ho M, *et al.* Effectiveness of precautions against droplets and contact in prevention of nosocomial transmission of severe acute respiratory syndrome (SARS). *Lancet* 2003;**361**:1519–20.

18. Lau JTF, Fung KS, Wong TW, Kim JH, Wong E, Chung S, *et al.* SARS transmission among hospital workers in Hong Kong. *Emerg Infect Dis* 2004;**10**:280–6.
19. Hui DS, Hall SD, Chan MTV, Chow BK, Tsou JY, Joynt GM, *et al.* Noninvasive positive-pressure ventilation. An experimental model to assess air and particle dispersion. *Chest* 2006;**130**:730–40.
20. Somogyi R, Vesely AE, Azami T, Preiss D, Fisher J, Fowler RA. Dispersal of respiratory droplets with open vs closed oxygen delivery masks: implications for transmission of severe acute respiratory syndrome. *Chest* 2004;**125**:1157.
21. Mardimae A, Slessarev M, Han J, Sasano H, Sasano N, Azami T, *et al.* Modified N95 mask delivers high inspired oxygen concentrations while effectively filtering aerosolized microparticles. *Ann Emerg Med* 2006;**48**:391–9.
22. Ip M, Tang JW, Hui DS, Wong ALN, Chan MTV, Joynt GM, *et al.* Airflow and droplet spreading around oxygen masks: a simulation model for infection control research. *Am J Infect Control* 2007;**35**:684–9.
23. Keady PB. Getting data you need with particle measurements. *Indoor Environ Connect* 2000;**2**:1–5.
24. Yu IT, Xie Z, Tsoi K. Why did outbreaks of severe acute respiratory syndrome occur in some hospital wards and not others? *Clin Infect Dis* 2007;**44**:1017–25.
25. Simonds AK. Continuous positive airway pressure and non-invasive ventilation in acute hypoxaemic respiratory failure. In Simonds AK, editor. *Non-invasive respiratory support: a practical handbook*. London: Hodder Arnold; 2007. pp. 57–71.